# Council Directive 98/57/EC of 20 July 1998 on the control of Ralstonia solanacearum (Smith) Yabuuchi et al.

# [F1 ANNEX II

## TEST SCHEME FOR DIAGNOSIS, DETECTION AND IDENTIFICATION OF *RALSTONIA SOLANACEARUM* (SMITH) YABUUCHI *ET AL.*

**Textual Amendments**

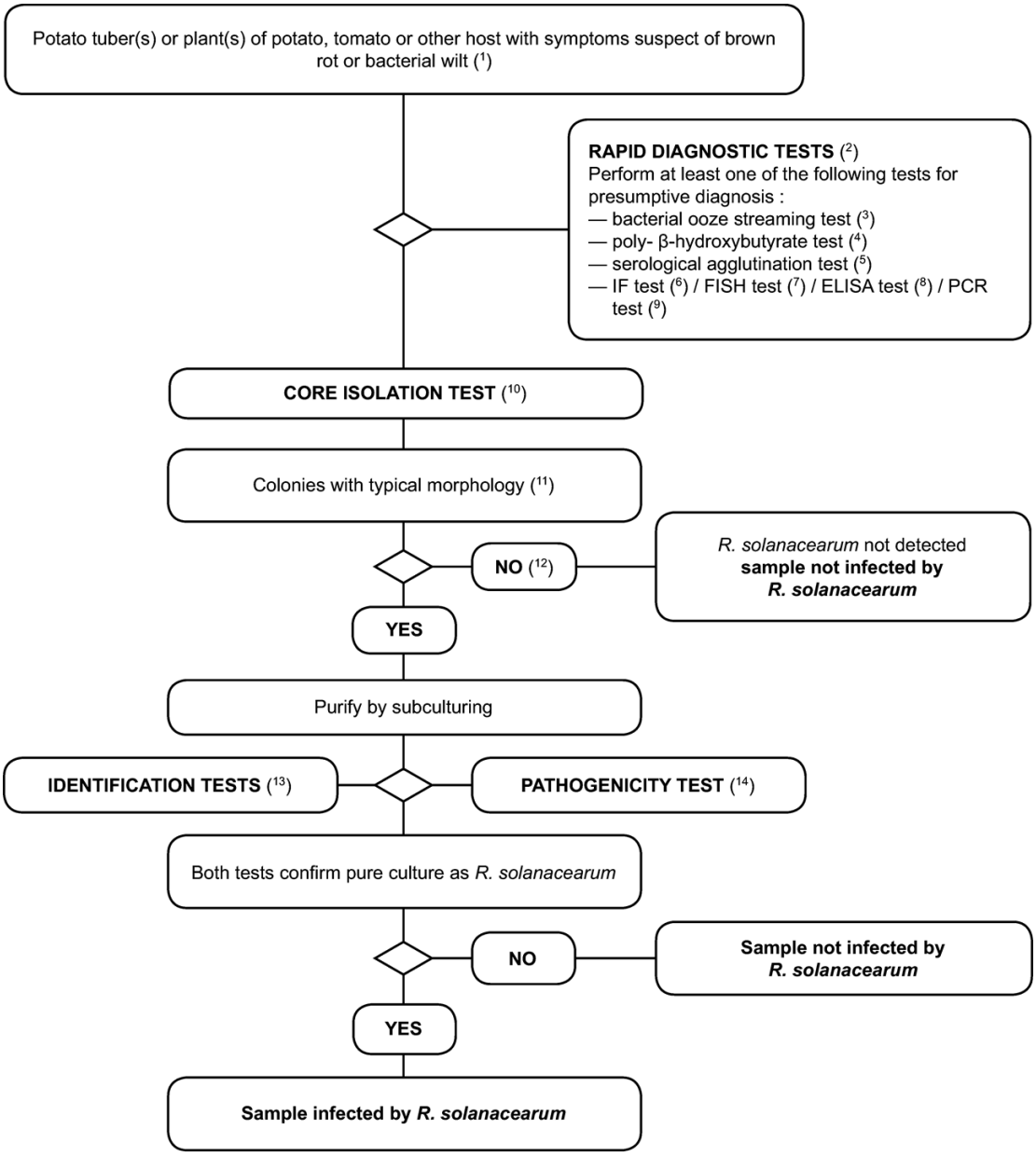
**F1** Substituted by Commission Directive 2006/63/CE of 14 July 2006 amending Annexes II to VII to Council Directive 98/57/EC on the control of Ralstonia solanacearum (Smith) Yabuuchi et al..

## SECTION I

## APPLICATION OF THE TEST SCHEME

1. Detection scheme for the diagnosis of brown rot and bacterial wilt (*Ralstonia solanacearum*) in potato tubers and potato, tomato or other host plants with symptoms of brown rot or bacterial wilt.

The testing procedure is intended for potato tubers and plants with symptoms typical or suspect of brown rot or vascular wilt. It involves a rapid screening test, isolation of the pathogen from infected vascular tissue on (selective) medium and, in case of a positive result, identification of the culture as *Ralstonia solanacearum*.]




(1) For description of symptoms see Section II.1.

(2) Rapid diagnostic tests facilitate presumptive diagnosis but are not essential. A negative result does not always guarantee absence of the pathogen.

(3) Streaming test for bacterial ooze from vascular stem tissue is described in Section VI.A.1.

(4) Test for poly-ß-hydroxybutyrate granules in bacterial cells is described in Section VI.A.2.

(5) Serological agglutination tests on bacterial ooze or extracts from symptomatic tissue are described in Section VI.A.3.

(6) IF test on bacterial ooze suspended in water or symptomatic tissue extracts is described in Section VI.A.5.

(7) FISH test on bacterial ooze suspended in water or symptomatic tissue extracts is described in Section VI.A.7.

(8) ELISA test on bacterial ooze suspended in water or symptomatic tissue extracts is described in Section VI.A.8.

(9) PCR test on bacterial ooze suspended in water or symptomatic tissue extracts is described in Section VI.A.6.

(10) The pathogen is usually easily isolated from symptomatic plant material by dilution plating (Section II.3).

(11) Typical colony morphology is described in Section II.3.d.

(12) Culturing may fail from advanced stages of infection due to competition or overgrowth by saprophytic bacteria. If disease symptoms are typical, but the isolation test is negative, then the isolation must be repeated, preferably using a selective plate test.

(13) Reliable identification of pure cultures of presumptive *R. solanacearum* isolates is achieved using the tests described in Section VI.B. Sub-specific characterisation is optional but recommended for each new case.

(14) The pathogenicity test is described in Section VI.C.